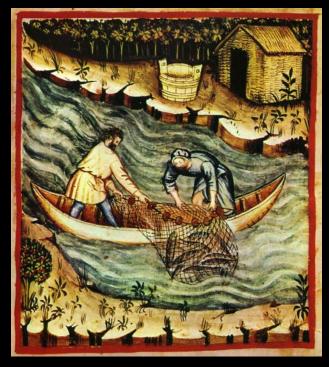
History of fisheries

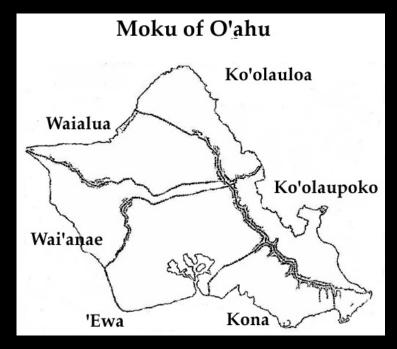


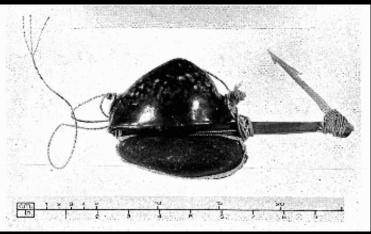


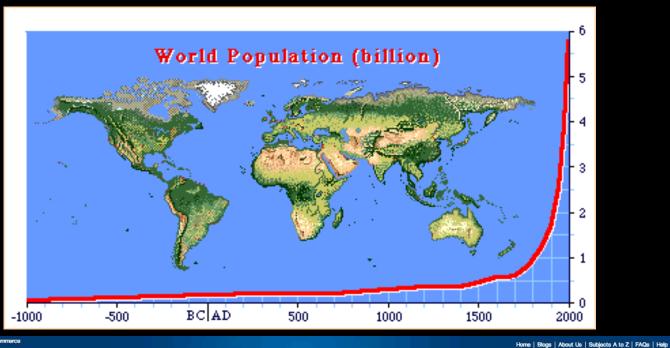


Local History and Traditions









Census

Business Geography D

Geography Data Research

Newsroom

Q Search Go

U.S. and World Population Clock

Note: The Population Clock is consistent with 2010 Census data and the most recent national population estimates.



Finfish and Invertebrates



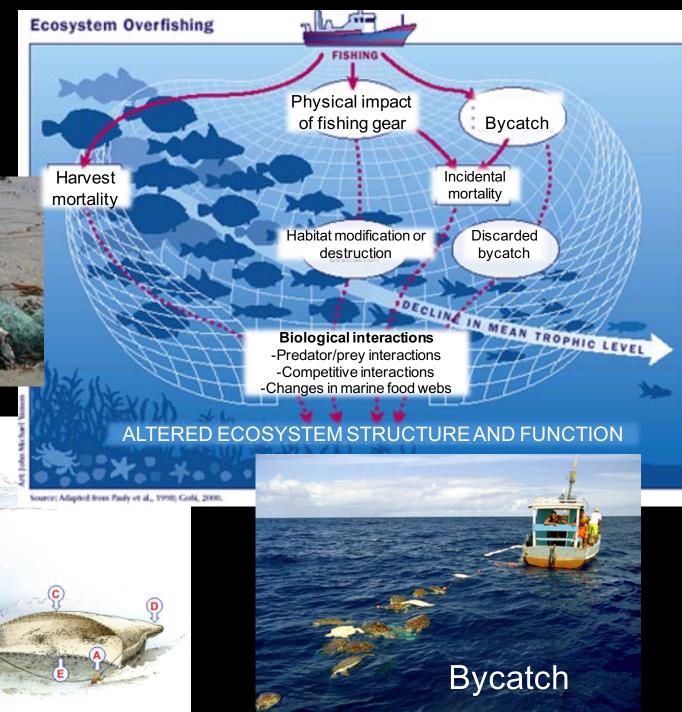






Overfishing

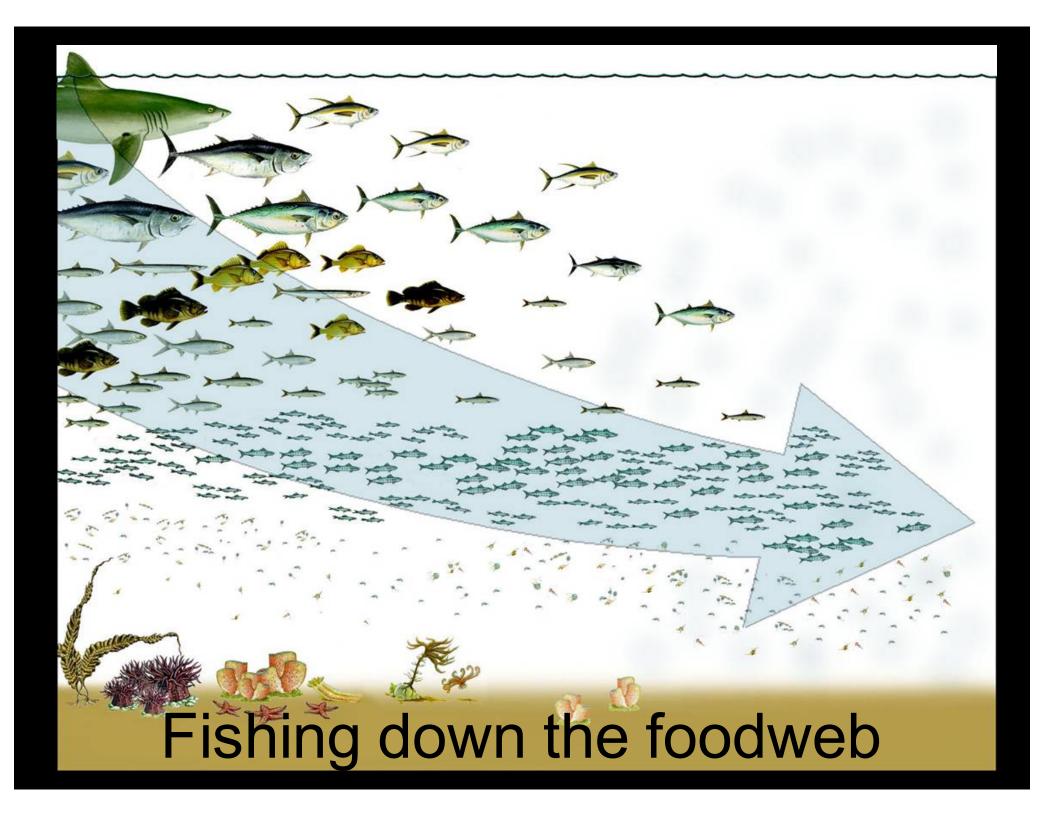
When fishing activities reduce fish stocks below a sustainable level.



Bottom trawler

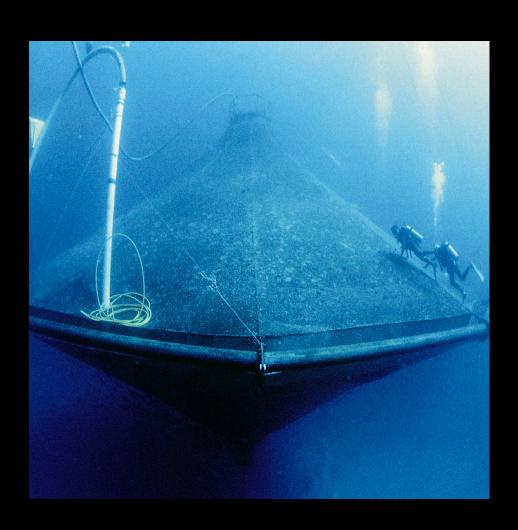
(CD012220-002)

Nets



There seem to be lots of problems...but are there any solutions?

ONE ALTERNATIVE: AQUACULTURE or FISH FARMING



ADVANTAGES:

- Reduces pressure on natural populations
- Reduces bycatch
- Reliable food source

DISADVANTAGES:

- Crowded cages can cause disease outbreaks
- Fish sewage and disease may escape and affect the environment
- Requires lots of research before fish can be raised and harvested

Hawaiian Fish Ponds: He'eia

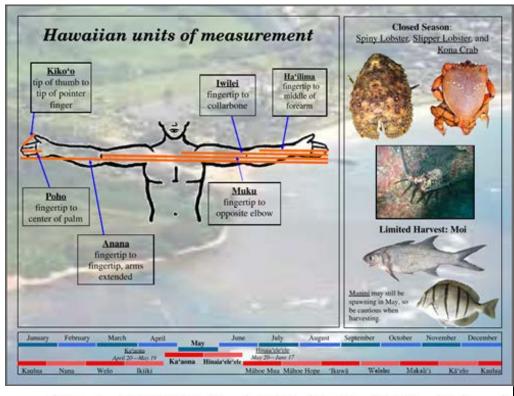
VIDEO

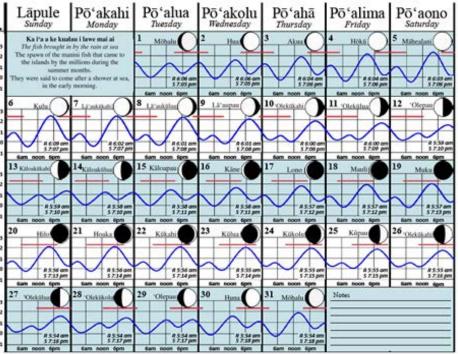


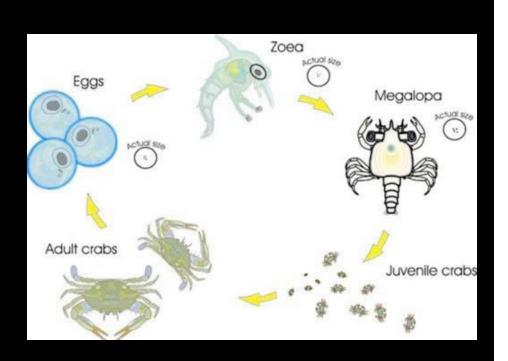
What do you know about fish spawning?

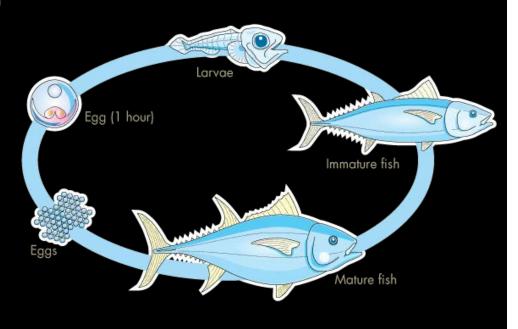
- Behavioral cues
- Environmental cues
- High mortality





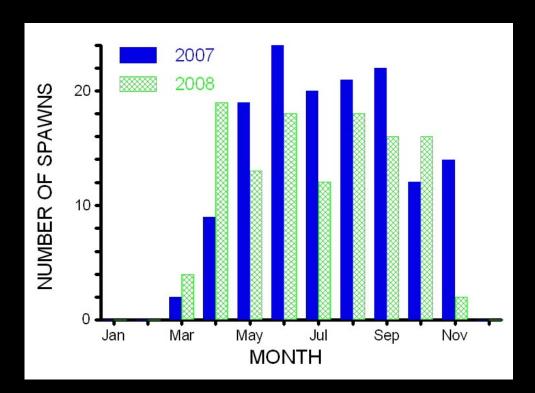






Coconut Island Research

Raising 'Ōpakapaka









Today's Activities

1. Collect spawn from 'Ōpakapaka pens



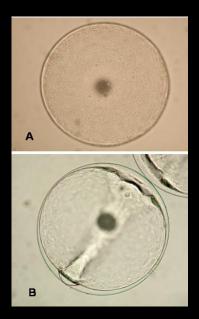
2. Quantify the number of eggs



3. Separate eggs from debris



4. Determine percent fertilization



5. Estimate time of spawning and hatching

COMPOUND MICROSCOPES



Task 2: Quantifying the number of eggs

- 1. Place aerator into bucket
- 2. Use glass pipette to mix first, and then transfer 10mL from bucket into petridish
- 3. Draw a grid and use magnifying glass + light to count eggs in 10mL
- 4. Calculate how many eggs are in 10L (multiply your count by 1000)
- 5. Take the average among all students in your group and report to instructors

Task 3: Separating eggs from debris

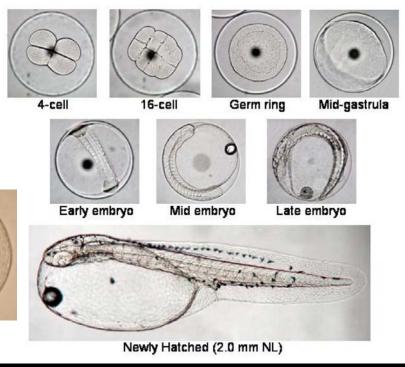
- 1. Turn off aerator
- 2. Use 1000mL glass beaker to remove 700mL from bucket FROM SURFACE
- 3. Add 300mL of tap water
- 4. Wait approximately 5 minutes for debris to surface
- 5. Use glass pipette to remove debris
- 6. Add Salt until the salinity matches the salinity of ocean water (USE REFRACTOMETER)
- 7. Wait approximately 5 minutes for eggs to surface
- 8. Use glass pipette to collect 10mL from the beaker (and as many eggs as possible) and transfer to little petri dish

1. Wait approximately 5 minutes for eggs to surface

2. Use glass pipette to collect 10mL from the beaker (and as many eggs as possible) and transfer to little petri dish

Task 4: Determine percent fertilization

- Use bulb pipette to transfer ~20 eggs from the beaker to a depression slide (watch demo)
- 2. Use compound microscope to how many eggs are fertilized
- 3. Calculate percent fertilization
- 4. Report findings to instructors



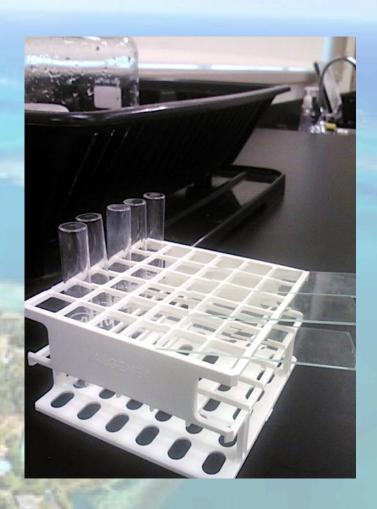
Task 5: Estimate time to spawning and time to hatching

- 1. Use Figure 10 to determine <u>developmental stage</u> of the eggs
- 2. Use Table in Appendix to calculate data and time of spawning and date and time of hatching
- 3. Report findings to instructors

CLEAN UP

- Wash all lab materials in the sinks
 - Clean pipettes by sucking up fresh water twice and rinse the outside.
- Use sink sponges to wipe down tables, then use blue/yellow rags and cleaner spray
- Sweep floors
- Place chairs on top of table





- Dry microscope slides and cover slips
- Dump water back in the ocean

RESULTS

- O How many eggs did each group find?
- What percentage of the eggs was fertilized?
 - What time did the spawning event occur?
 - When can the eggs be expected to hatch?